



# *X-Ray Damage to Biological Crystalline Samples*

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# *International Workshops on X-Ray Damage to Biological Crystalline Samples*



First Workshop (RD1) 1999 at ESRF

Second Workshop (RD2) 2001 at APS

<http://www.aps.anl.gov/News/Conferences/2001/damage/home.html>

8 papers in Journal of Synchrotron Radiation (2002) **9**, 327 - 382

Third Workshop (RD3) 2003 at ESRF

9 papers in Journal of Synchrotron Radiation (2005) **12**, 257 - 329

Fourth Workshop (RD4) 2006 at SPring8

<http://www.spring8.or.jp/en/users/meeting/rd4>

several presentations will be published

Organizers:

Elspeth Garman, Collin Nave, Gerd Rosenbaum, Raimond Ravelli, Seam McSweeney



# *Radiation Damage - An Unavoidable Byproduct Of Crystallographic Data Acquisition*

- Radiation damage is proportional to dose on sample.

dose = absorbed energy / mass ( unit: 1 Gy = 1 J/kg )

dose =  $\mu / \rho I t \epsilon_{\text{phot}}$  I = flux density, t = exp. time

- Number of photons in diffraction peak

$N_{\text{phot}} \sim I t V \lambda^2$  V = sample volume

- Dose on sample / number of photons in diffraction peak

dose /  $N_{\text{phot}} \sim 1/V$

Almost all deposited energy is by photoelectric absorption.

$(\mu / \rho)_{\text{photo}} \sim \lambda^3 \quad \Rightarrow \quad \text{dose} / N_{\text{phot}} \sim \lambda^2 / \lambda^2$

- Radiation damage to cryo-cooled samples is **not** a particular property of 3<sup>rd</sup> gen. sources.

Lavish use of available flux **is** characteristic for users of 3<sup>rd</sup> gen. sources: over-exposure

Detector read-out units should be calibrated in photons to help guide level of exposure.



# *Primary and Secondary Radiation Damage and Cryo-cooling*

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## Primary radiation damage:

Direct hit of protein by absorbed photon or by ejected photo-electron

## Secondary radiation damage:

Damage of protein by action of non-protein molecules activated by absorbed photon or by photo-electron (mostly hydroxyl radicals)

## Data Collection at Room Temperature:

Early observation of radiation damage on home sources.

More than 1 sample per data set needed.

## Cryo-cooling of samples:

Radicals immobilized at 100 K; secondary damage stopped;

~50-100-times increase in sample life (dose tolerance);  $I_{1/2}$  dose =  $4 \cdot 10^7$  Gy

Cooling to 15 K: improvement not clearly demonstrated; B-factor reduced (mostly)



## *Radiation Damage Observed at 3rd Generation Sources*

### Renewed Attention to Radiation Damage at 3rd Generation Sources:

- observed higher radiation damage attributed to high flux density
- no significant dose rate effect observed in carefully designed studies
- photons from 3rd gen. sources are not different from photons from other sources
- high flux allows much higher exposure in available beam time allotment
- concern about beam heating:
  - not the cause of increased damage, even at max. flux from APS undulator
  - temperature increase <10 K

### X-Ray Damage is Not Local

- photo-electrons carry ~95% of energy of absorbed photon (12 keV)
- photo-electrons from 12 keV photons travel ~2.8  $\mu\text{m}$
- transfer damage energy of 20 eV average per hit
- only 5% of energy (binding energy of C, N, O) of absorbed photon stays within 5  $\text{\AA}$





# *Effects of Radiation Damage*

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Breaking bonds => loss of occupancy, increased temp. factor  
S-S bonds first, then carboxyl, then other charged residues, then ...  
(It's the chemistry, not the local absorption cross-section.)

Changing charge state

Reduction of metal centers

Ironically, cryo-protectant among worst promoters.

Increase in unit cell volume and lattice constants

General decrease of intensity of diffraction peaks

Increase in B-factor

Non-Isomorphism

Increased  $R_{\text{merge}}$  between initial and later frames

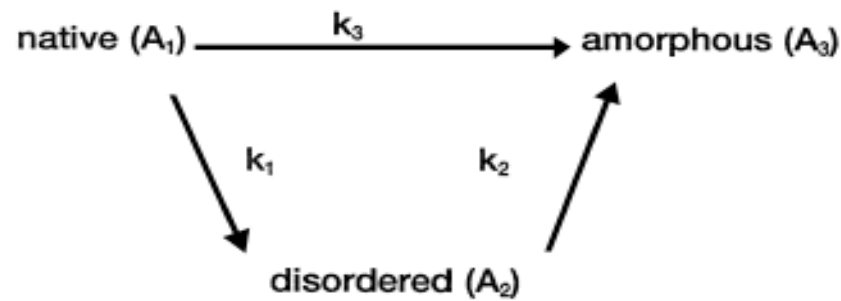
Problem for SAD/MAD phasing, especially sulfur SAD

(Byfoet pairs suffered different amount of radiation damage)



# Process of Radiation Damage

Blake and Phillips [1] suggest the following model for radiation damage :



The “disordered” unit cells have elevated temperature factors; the amorphous regions do not give Bragg diffraction at all. Hendrickson [2] concluded from data collected from myoglobin at room temperature that  $k_3=0$ . Sliz et al. [3] concluded from data collected from several samples at 100 K at much higher exposure that  $k_3=0$ .

The process of radiation damage is sequential:  $A_1 \Rightarrow A_2 \Rightarrow A_3$

1. Blake, C.C. & Phillips, D.C. (1962). Effects of X-irradiation on single crystals of myoglobin. In *Biological Effects of Ionizing Radiation at the Molecular Level*. (Vienna: International Atomic Energy Agency), pp. 183-191.
2. Hendrickson, W.A. (1976). Radiation damage in protein crystallography. *J. Mol. Biol.* 106, 889-893.
3. Sliz, P., Harrison, S.C. & Rosenbaum, G. (2003). How does Radiation Damage in Protein Crystals Depend on X-Ray Dose. *Structure* 11, 13-19



# *Damage vs. Dose, Dose Rate Minimum Sample Size*

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Studies of damage vs. dose and limiting sample sizes by Teng & Moffat (2000, 2002), Glaser et al. (2000) and Sliz, Harrison, Rosenbaum (2003)

Radiation damage proportional to dose up to a certain limit; then fast collapse of structure (non-linear dose effect, not dose rate effect)

No dose rate effect noticeable for flux densities up to  $3 \cdot 10^{15}$  ph/s/mm<sup>2</sup>

T&M and S,H,R conclude damage in cryo-cooled samples is primary damage, i.e. radicals are immobilized.

Limit of crystal volume for full data set:

Glaser et al.:  $\sim 35 \mu\text{m}$

Teng&Moffat:  $\sim 30 \mu\text{m}$

Sliz, Harrison, Rosenbaum:  $\sim 15 \mu\text{m}$

(different resolutions, different criteria for damage limit.)





# Typical Effects of Radiation Damage on Diffraction Intensities, $R_{merge}$ , Unit Cell Volume, B-factor

Fig 1

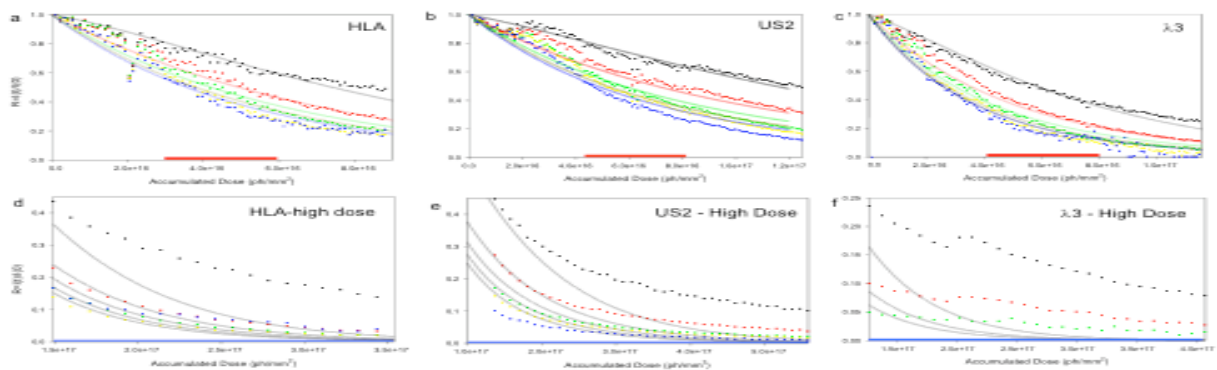
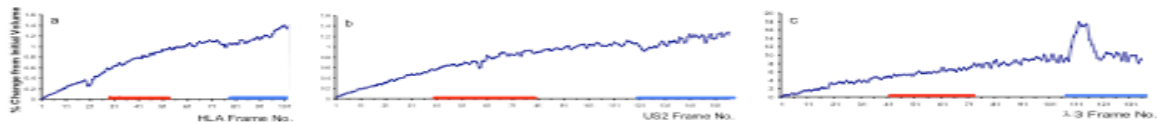


Fig 2



Fig 3



# *Preventing, mitigating the effects of radiation damage*

- **Minimize dose**
  - avoid high Z atoms in buffer
  - calculate dose using program RADDDOSE [1]
  - reduce exposure to level necessary for accuracy as determined by photon statistics
  - reduce scatter by
    - minimizing beam on non-crystal volume
    - minimizing path length of primary beam through air
    - clean freeze, no ice
    - avoid high Z atoms in buffer (e.g. arsenic) or other high mol. weight stuff
- **Radio-protectants (Garman, others)**

Scavengers: effectiveness not clear; concern about chemistry

1. Murray et al., J.Appl.Cryst. **37**, 513-522  
(to get program contact Elspeth Garman [elspeth@biop.ox.ac.uk](mailto:elspeth@biop.ox.ac.uk))



## *Mitigating or make use of the effects of radiation damage*

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- Correction of diffraction peak intensities for radiation damage (Otwinowsky, others)  
Akin to dead time loss correction. Extrapolate back to zero exposure
- Use non-isomorphism for phasing (Ravelli, others)
- Technology developments that can help reducing dose
  - detectors with low read noise / signal per photon)
  - ratio (read noise / signal per photon) should be  $<1$
  - detectors with large dynamic range  
(max. no. of photons per pixel / read noise in photons)

